

A scanning electron micrograph (SEM) showing a complex biofilm structure. The biofilm consists of numerous rod-shaped bacterial cells, some of which are covered in a dense, textured layer of purple material, likely representing extracellular polymeric substances (EPS) or a protective matrix. The cells are interconnected and form a three-dimensional network. The background is a dark, slightly textured surface.

# Challenges in evaluating high-level disinfectants against biofilms

CARDIFF  
UNIVERSITY

PRIFYSGOL  
CAERDYDD

Prof. Jean-Yves Maillard (Maillardj@Cardiff.ac.uk)

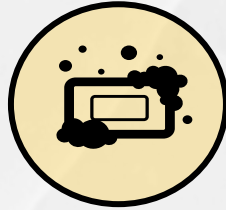
# Transparency declaration

**Director of Biocide Consult Ltd**

**Chief Editor, Journal of Infection Prevention**

– nothing pertinent to this presentation to declare

# Cleaning vs. disinfection



## CLEANING

- Removal of dirt
- Not designed to kill pathogens
- Might (and probably will) remove pathogens



## DISINFECTION

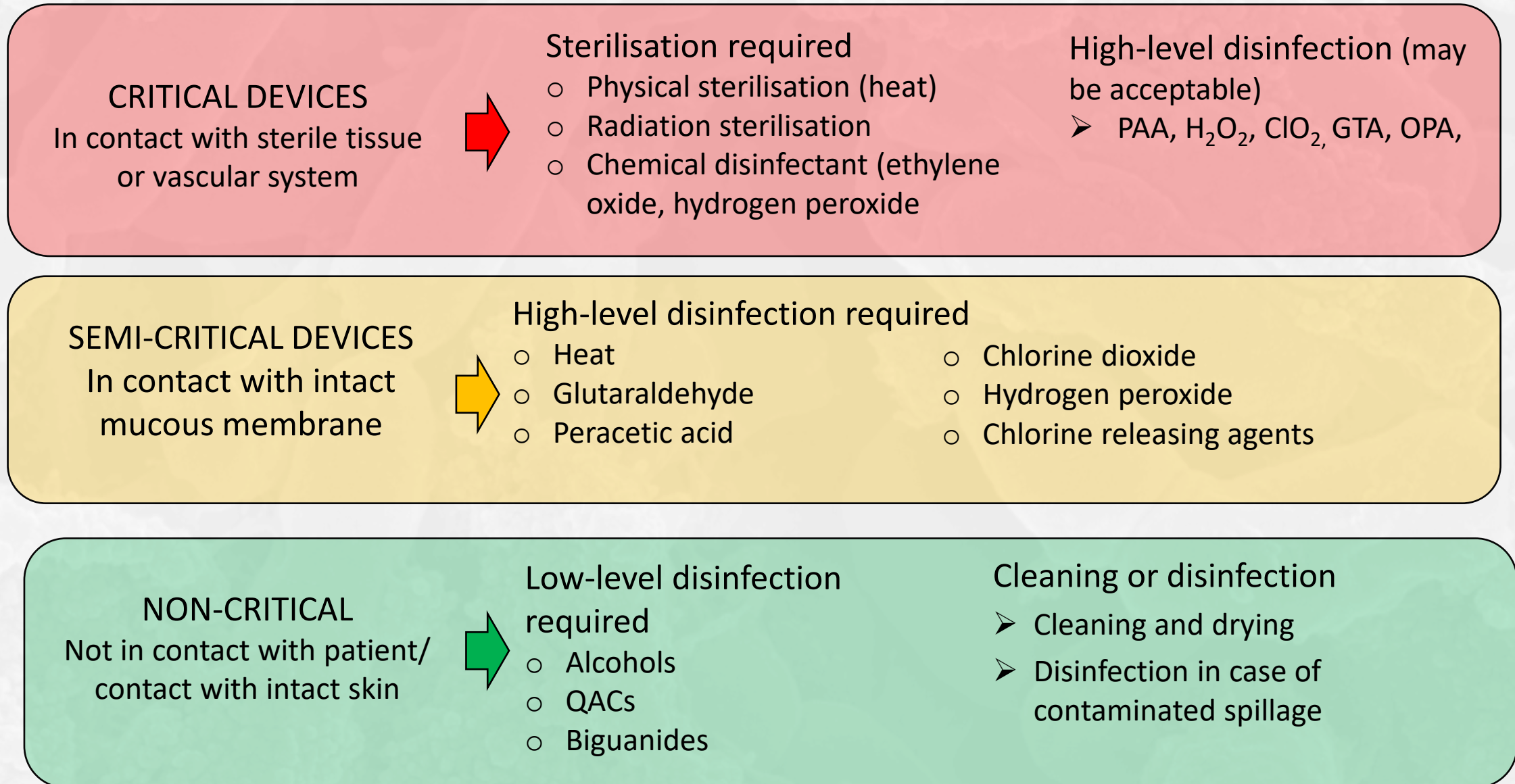
- Killing pathogens – a “cidal” activity
- Different levels of disinfection
- Delivery (applicator) plays a role



It is not possible to ‘sterilise’ surfaces of equipment/ patient environment using disinfection; only to temporarily reduce the number of microbes present.

# Revised Spaulding Classification

Spaulding classification



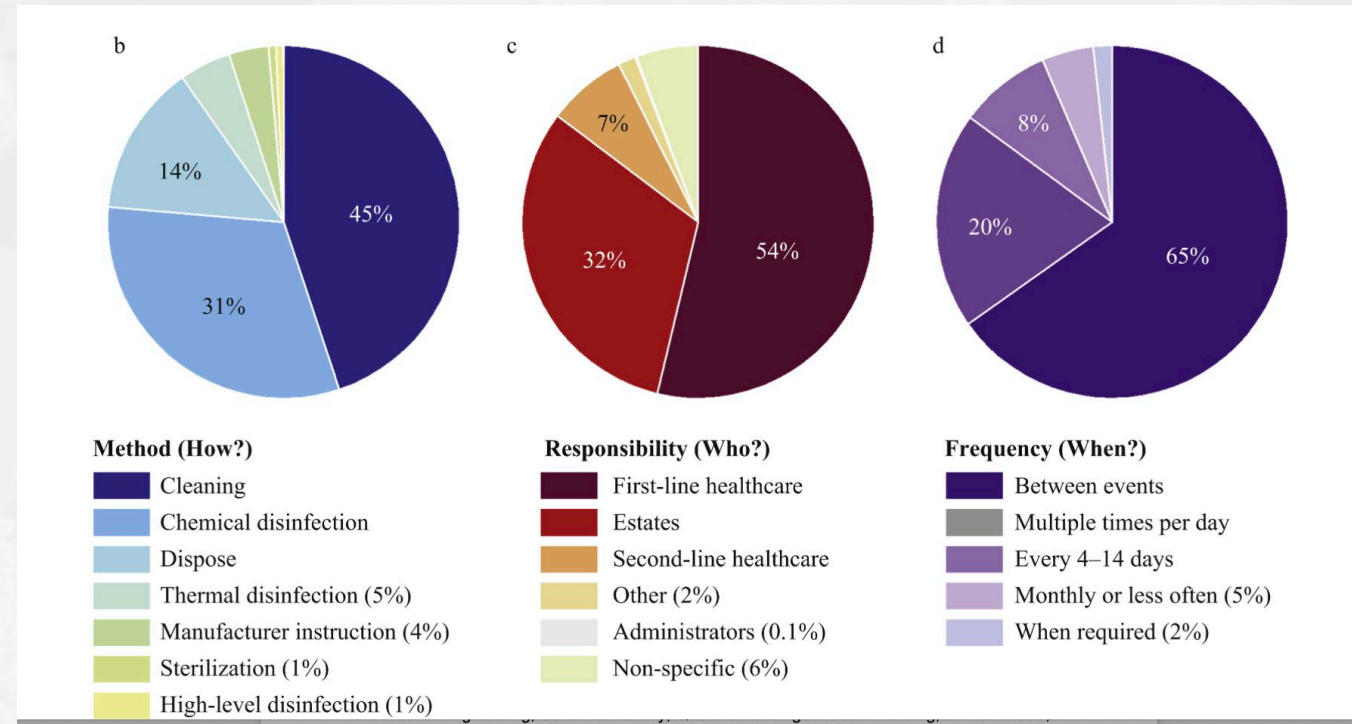
# Decontamination protocols for shared non-critical objects

➤ The lack of clear indications regarding the person responsible for the decontamination process put at risk the ability of policies to serve as guidance

Castelli *et al.* *J Hosp Infect* 2022;120:65-72.

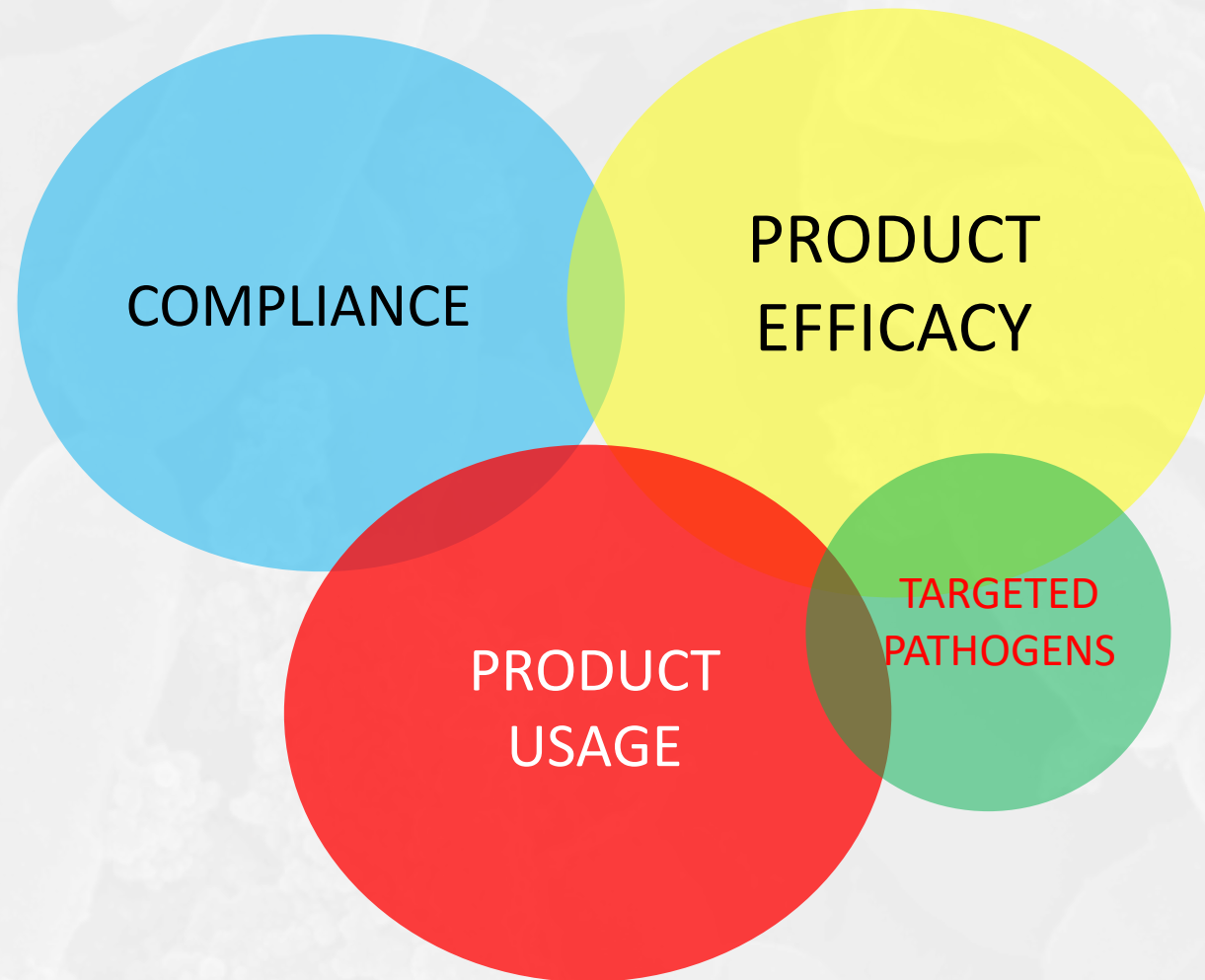
- 35 NHS Acute Care Organisations
- 1279 decontamination protocols regarding 283 different shared non-critical objects were retrieved.

- 689 (54%) did not indicate the person responsible for decontamination
- Only 425 (33%) were complete, giving indications for (a) decontamination method, (b) responsibility and (c) frequency
- Only 2.5% (32/1279) of decontamination protocols were complete and identical in two policies



Summarized indications retrieved for the decontamination method (b), responsibility (c) and frequency (d).

# Factors impacting Disinfectant efficacy



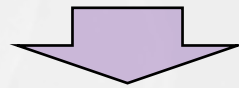
# Factors impacting Disinfectant efficacy

## FACTORS INHERENT TO THE PRODUCT



- Concentration
- Formulation
- Delivery system

## FACTORS INHERENT TO THE APPLICATION OF THE PRODUCT



- Surface
- Organic load (soiling)
- Contact time
- Temperature
- Humidity

## FACTORS INHERENT TO THE MICRO-ORGANISMS



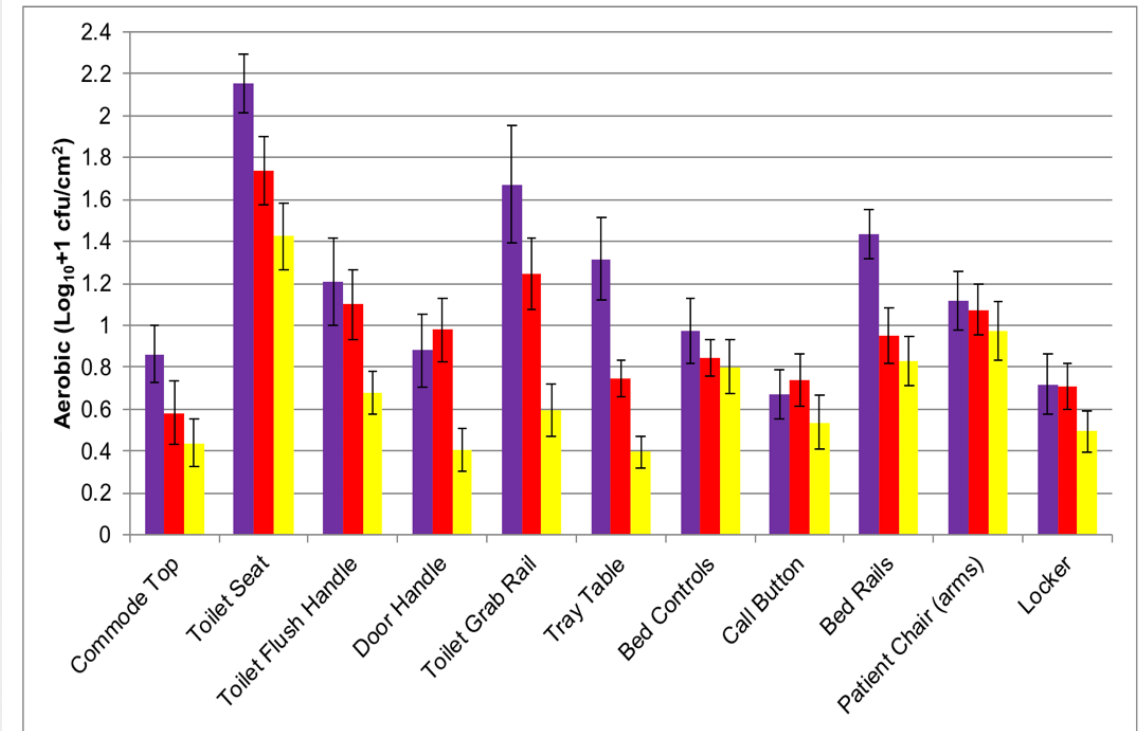
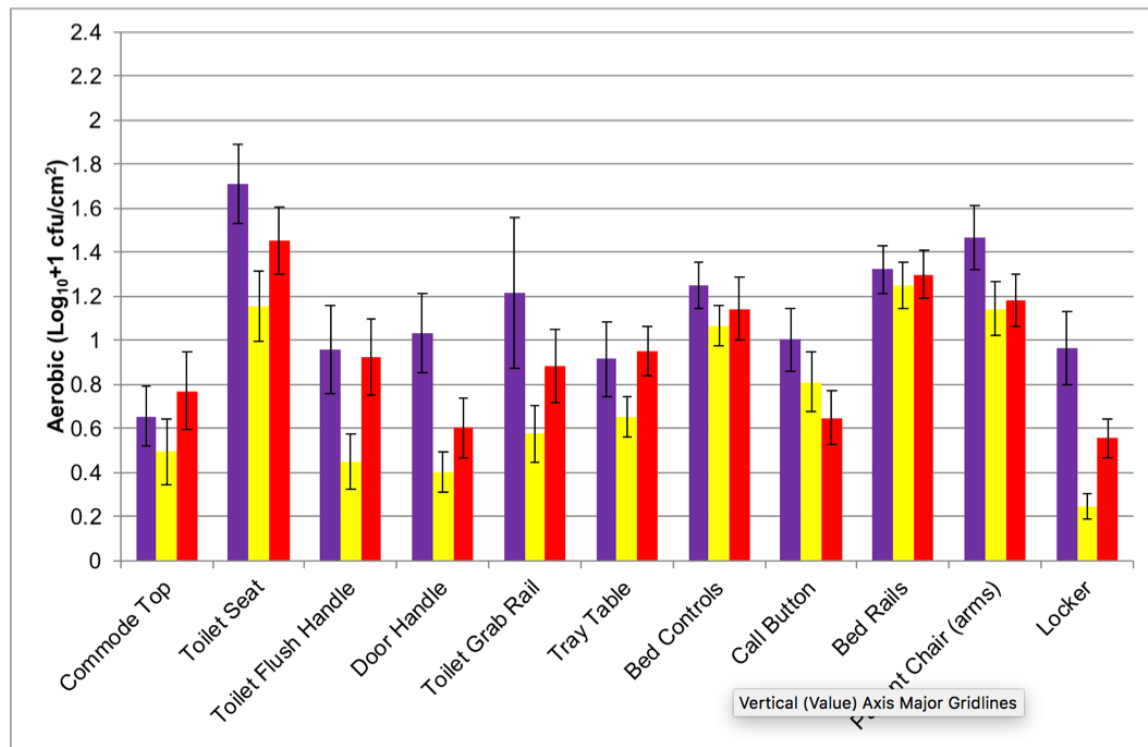
- Type
- Number
- Association (biofilms)

# Factors impacting Disinfectant efficacy

Delivery – preformulated wipe vs. bucket & cloth

Siani *et al. Am J Infect Control* 2018 46;1180-7

Double-crossover study was performed on 2 different surgical and cardiovascular wards in a 1,000-bed teaching hospital over 29 weeks.



- Baseline
- Sporicidal wipe
- Cleaning and use of chlorine 1,000 ppm (bucket)

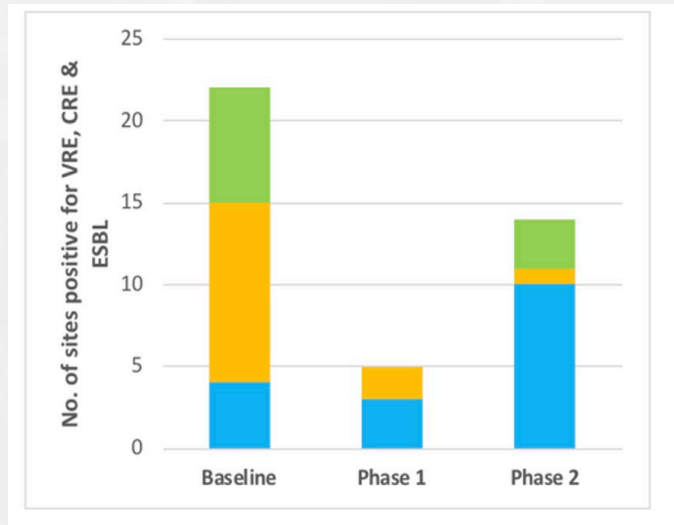
- Baseline
- Cleaning and use of chlorine 1,000 ppm (bucket)
- Sporicidal wipe

# Factors impacting Disinfectant efficacy

Delivery – preformulated wipe vs. bucket & cloth

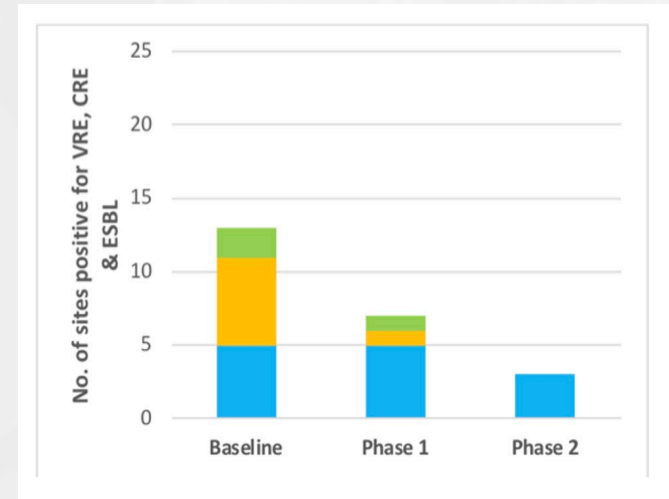
Siani *et al.* *Am J Infect Control* 2018 46;1180-7

Double-crossover study was performed on 2 different surgical and cardiovascular wards in a 1,000-bed teaching hospital over 29 weeks.



## Ward 1

- Phase 1: Sporicidal wipe
- Phase 2: Cleaning and use of chlorine 1,000 ppm (bucket)



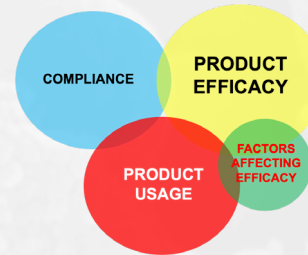
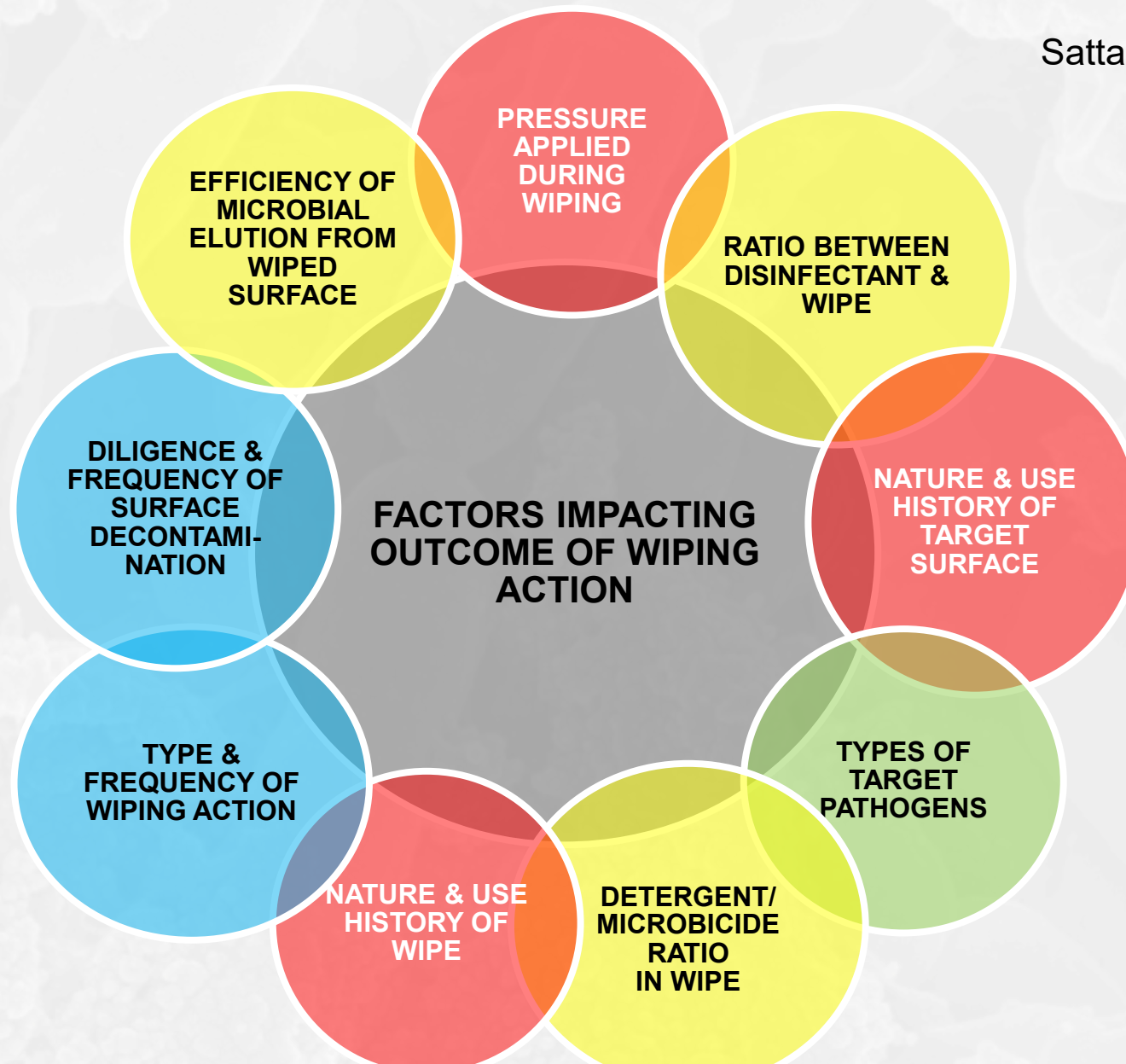
## Ward 2

- Phase 1: Cleaning and use of chlorine 1,000 ppm (bucket)
- Phase 2: Sporicidal wipe

- Extended-spectrum beta lactamases Enterobacteriaceae (ESBL)
- Carbapenem-resistant Enterobacteriaceae (CRE)
- Vancomycin-resistant enterococci (VRE).

# Factors impacting Disinfectant efficacy

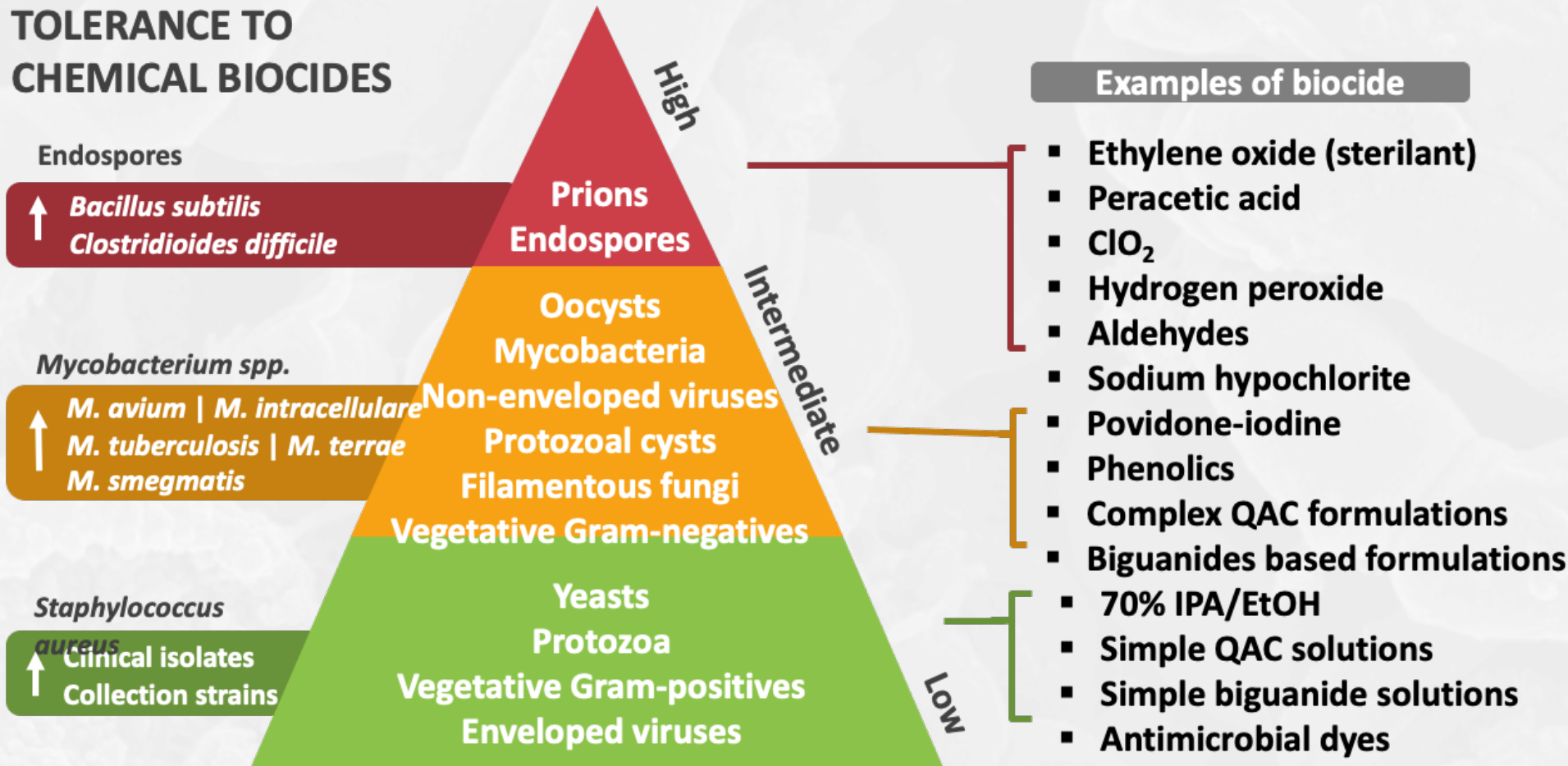
Sattar & Maillard *Am J Infect Control* 2013 41;S97-S104



# Factors impacting Disinfectant efficacy

Maillard & Pascoe. *Nature Rev Microbiol* 2023: doi.org/10.1038/s41579-023-00958-3

## TOLERANCE TO CHEMICAL BIOCIDES



# Factors impacting Disinfectant efficacy

## SPORICIDAL ACTIVITY

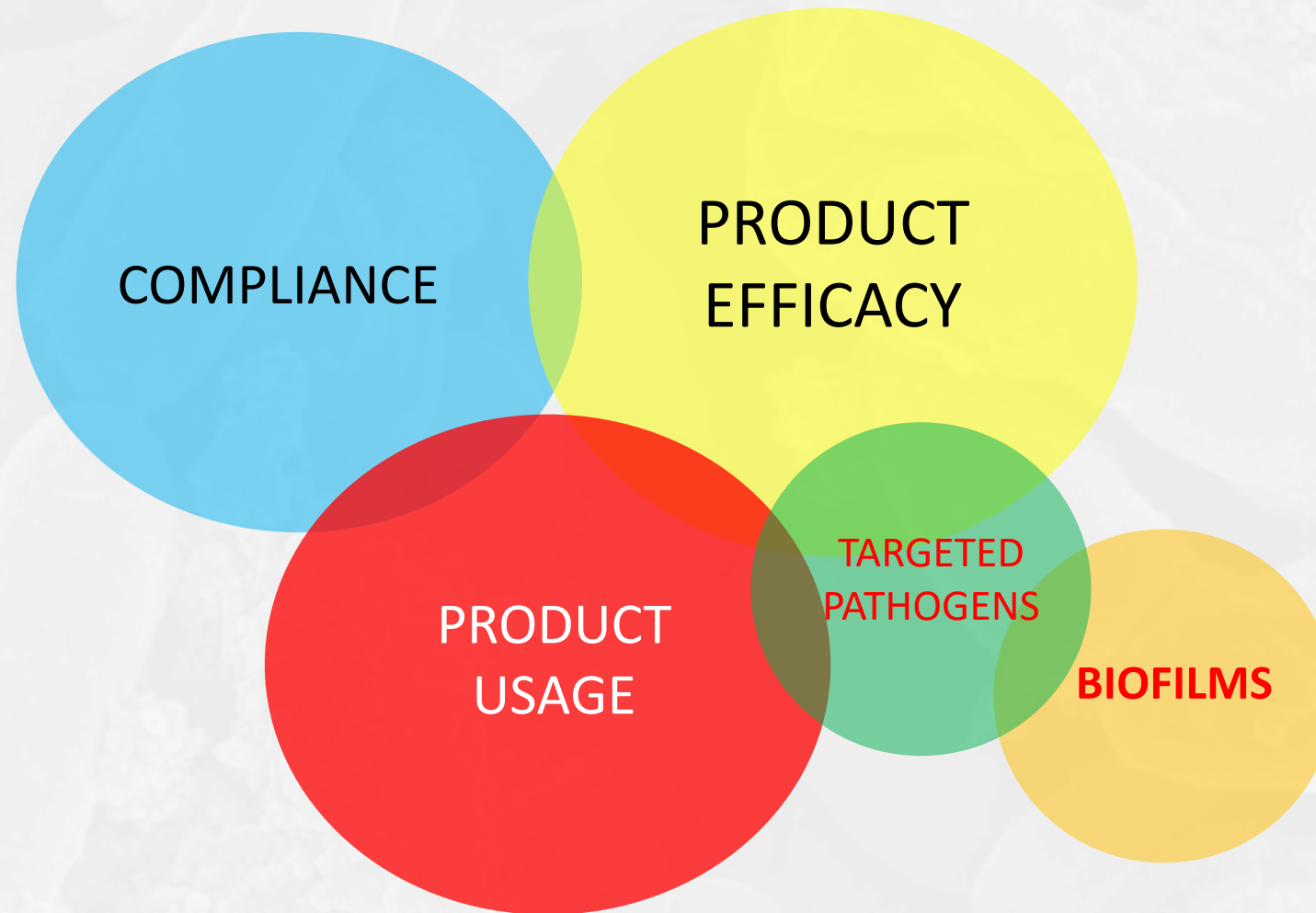
- Hydrogen peroxide
- Peracetic acid
- Chlorine dioxide
- Ozone
- Ethylene oxide
- Glutaraldehyde
- Formaldehyde
- *ortho*-phthalaldehyde
- Sodium hypochlorite
- Sodium dichlororiscyanurate (?)
- Chloramine-T
- Calcium hypochlorite
- Iodine and iodophors

Russell AD. Clin Microbiol Rev 1990;3:99-119.

## NOT SPORICIDAL

- Phenols and cresols
- Quaternary ammonium compounds
- Biguanides
- Organic acids and esters
- Alcohols

# Factors impacting Disinfectant efficacy



# Importance of biofilm in healthcare settings



- 137 healthcare professionals completed the online survey
- 87.6% of all participants had heard of the term biofilm
- 83.9% knew what the term biofilm means.
  - Medical device (86.1%)
  - Drain biofilm (76.5%)
  - 39.1% knew about dry surface biofilms



Isabella  
Centeleghe

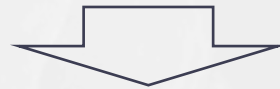
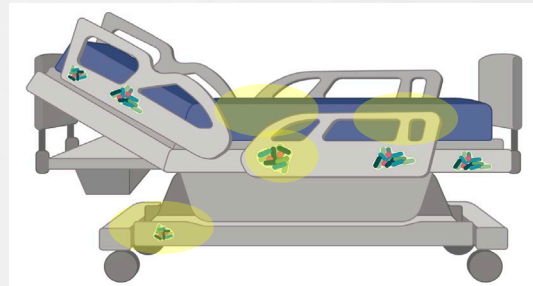
# Biofilms-related parameters to consider

## Drain Biofilms



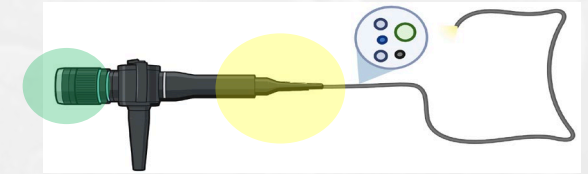
- Reduced microbicidal effect
- Decreasing biofilm biomass
- Regrowth post-intervention

## Dry Surface Biofilms



- Reduced microbicidal effect
- Preventing transfer – ensuring the surface is safe
- Detection

## Medical device biofilms



- Effective elimination of all microorganisms in all the medical device parts
- Regrowth post-intervention

# Importance of biofilm in healthcare settings

Adapted from Otter *et al. J Hosp Infect* 2015; 89:16e27

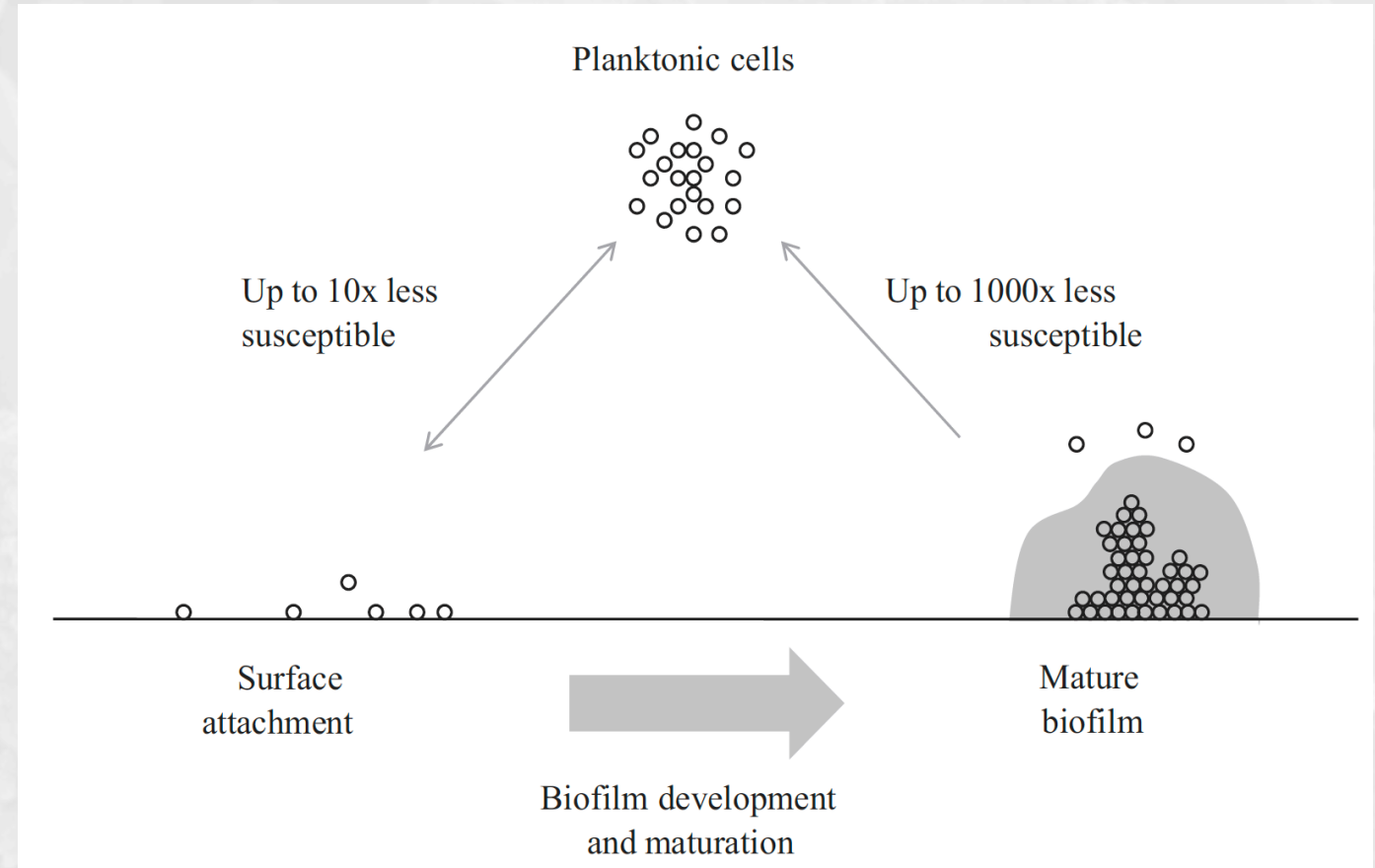
## Susceptibility to disinfection

Planktonic suspension

Planktonic dried on surfaces

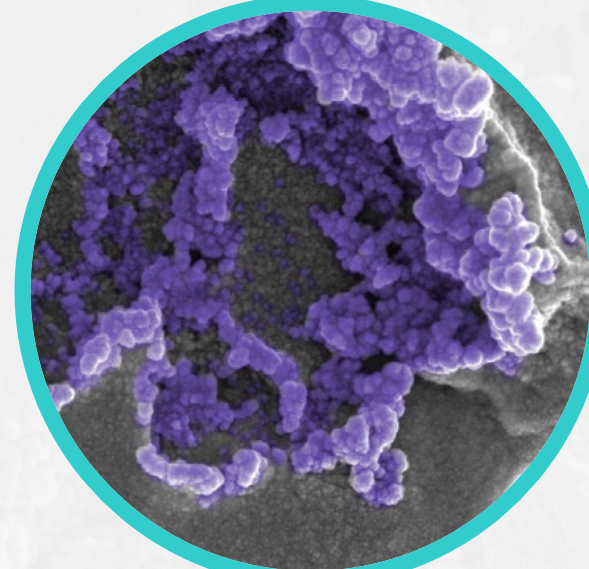
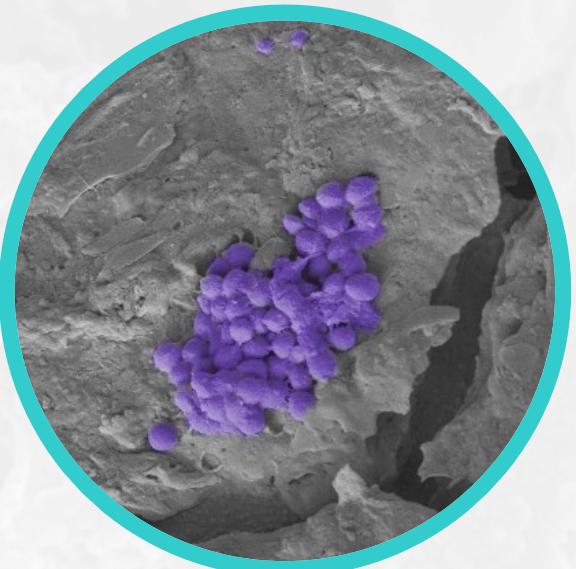
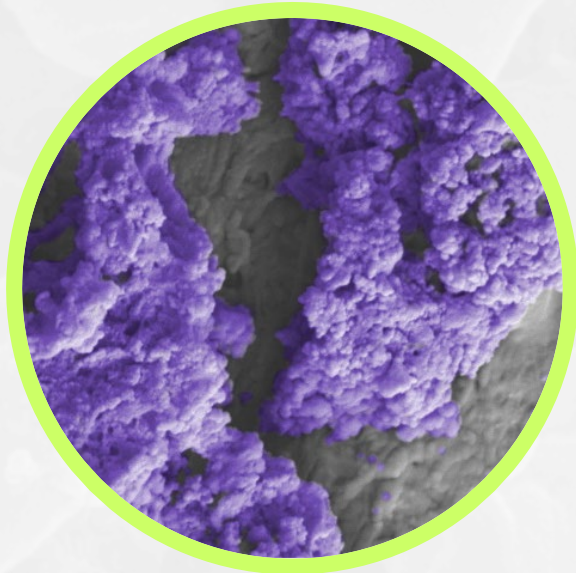
Hydrated biofilms

Dry surface biofilms



# DSB in healthcare settings

Ledwoch et al. J Hosp Infect 2018;100:3:e47-e56



- Dry surface biofilms are **widespread** on surfaces in hospitals (**90-95 % surfaces sampled**)
- DSB contribute to pathogens survival **despite cleaning and disinfection**
- DSB **cannot be detected** by swabbing or contact plates when surfaces are dried

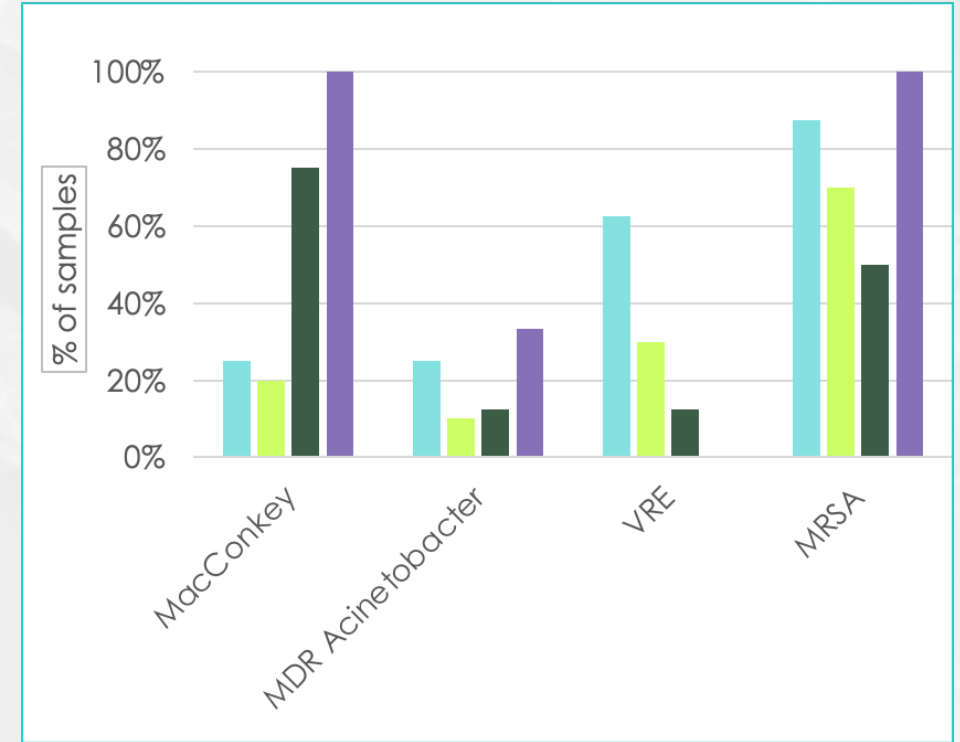
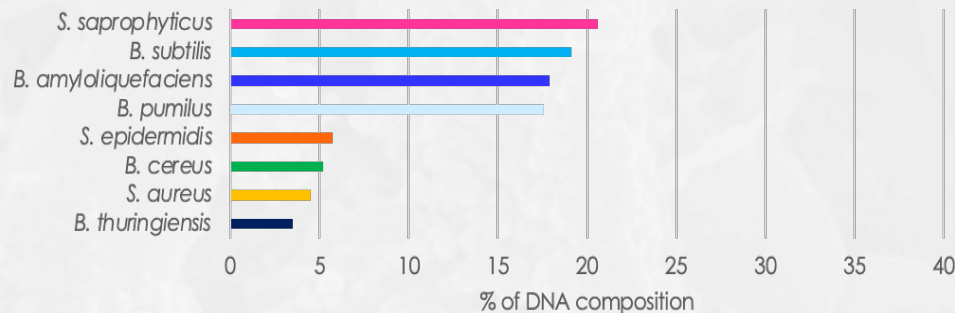
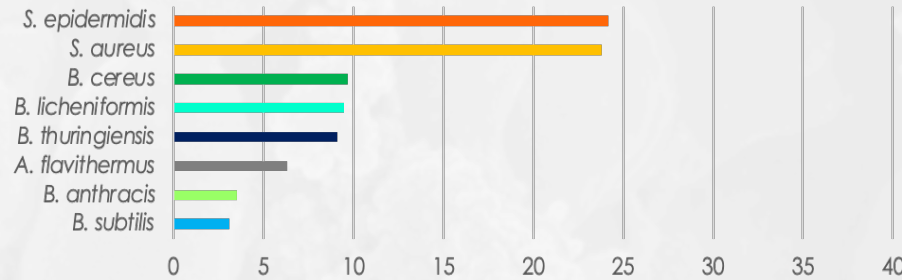
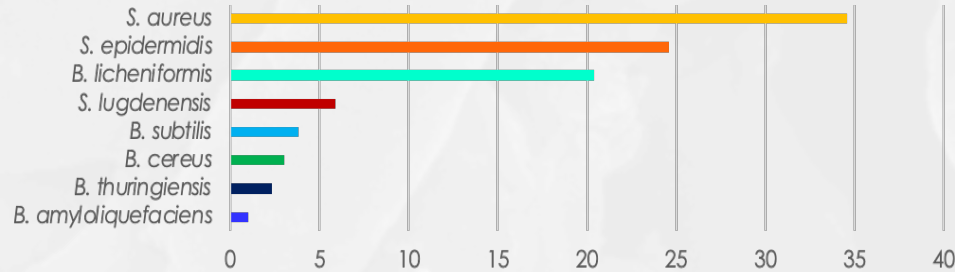
# DSB composition

Ledwoch et al. J Hosp Infect 2018;100:3:e47-e56

Ledwoch et al. J Hosp Infect 2021;112:3:31-6

## Next Generation Sequencing (16s rRNA)

## Selective agar



Keyboard samples from:  
 ■ WH - Welsh Hospital  
 ■ SH - Scottish Hospital  
 ■ EH - English Hospital  
 ■ SD - Scottish Dental practice

# DSB in healthcare settings

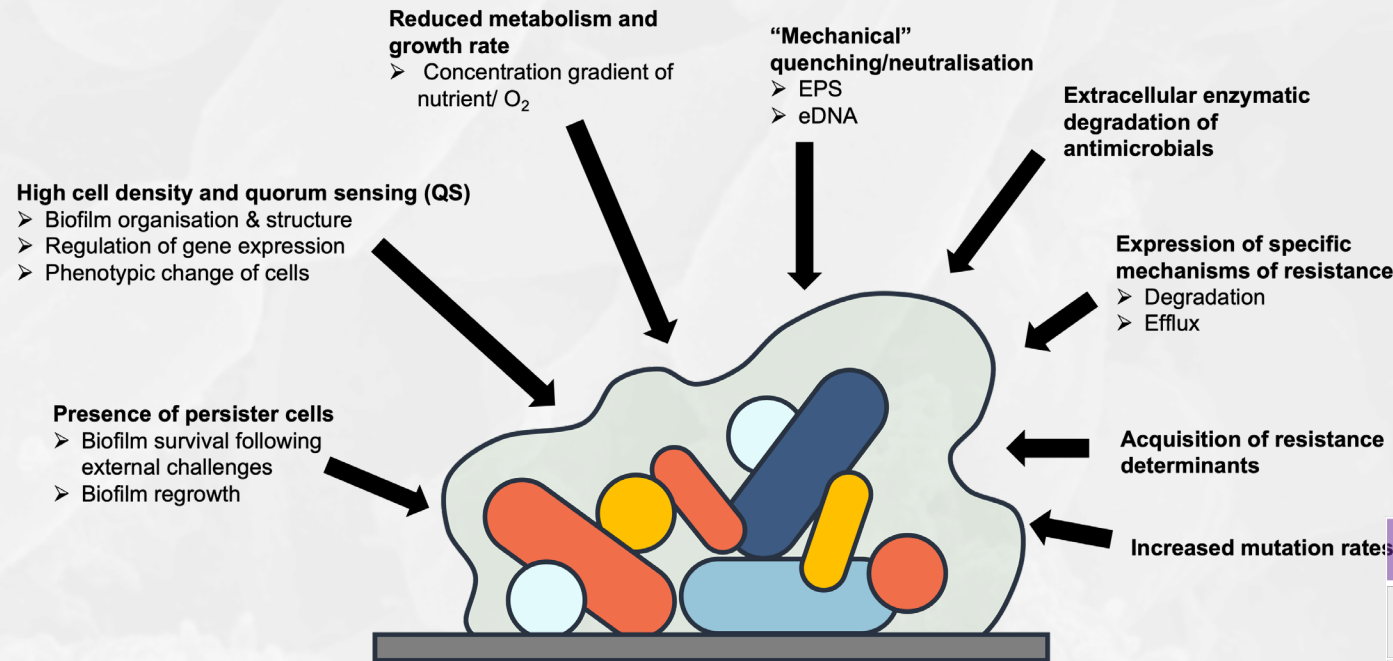
Ledwoch et al. J Hosp Infect 2021;112:31-36.

Keyboard sample number	Origin	Healthcare facility	Bacteria from DSB detected (+) / not detected (-)	
			Transfer test after wiping with sterile water <sup>2</sup>	Transfer test after wiping with NaOCl 1,000 ppm <sup>2</sup>
1	Wales	1,000-bed hospital	+	+
2			+	-
3			-	-
4			-	-
5	Scotland	500-bed hospital	+	-
6			+	+
7			-	+
8			+	-
9	England	1,700-bed hospital	-	+
10			+	+
11			+	-
12	Scotland	Dental practice	+	+
13			+	+
Total			9/13	7/13

- Pathogens from **69% of samples could be transferred** following wet wiping with sterile water.
- **54% of samples continued to transfer bacteria** following 1,000 ppm NaOCl treatment.

# Dry surface biofilm vs. hydrated biofilms

Maillard & Centeleghe. *Antimicrob Res Infect Control* 2023;12:95.



	DSB	Hydrated biofilms
EPS	✓	✓
eDNA	?	✓
Depth (gradients)	✗	✓
Desiccation	✓	✗
Reduced metabolism	✓	✓
Persister cells	?	✓

# Efficacy of disinfectants against hydrated biofilms

## Hydrated biofilms

- Complex microbial communities formed and grown in wet /semi-wet habitats.
- Colonize various surfaces (drain, medical equipment)
- Clean + mechanical action + disinfection

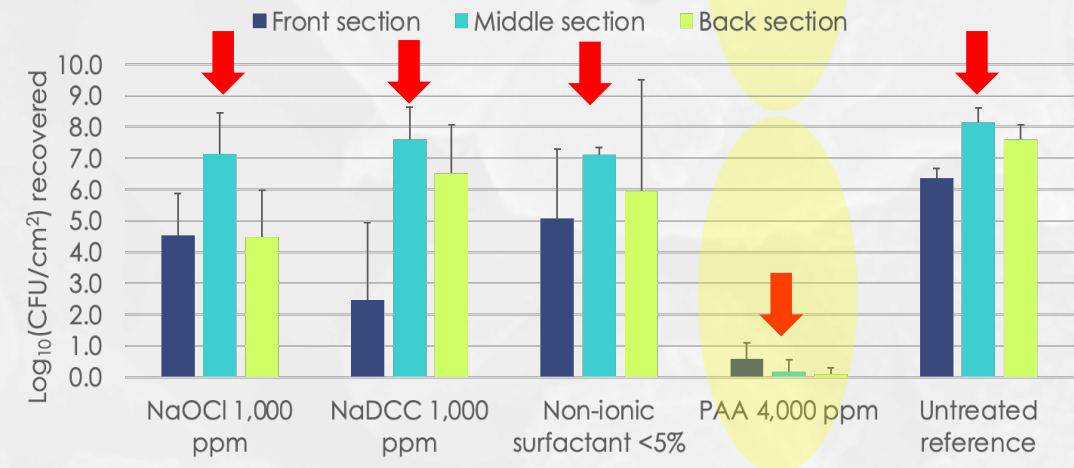
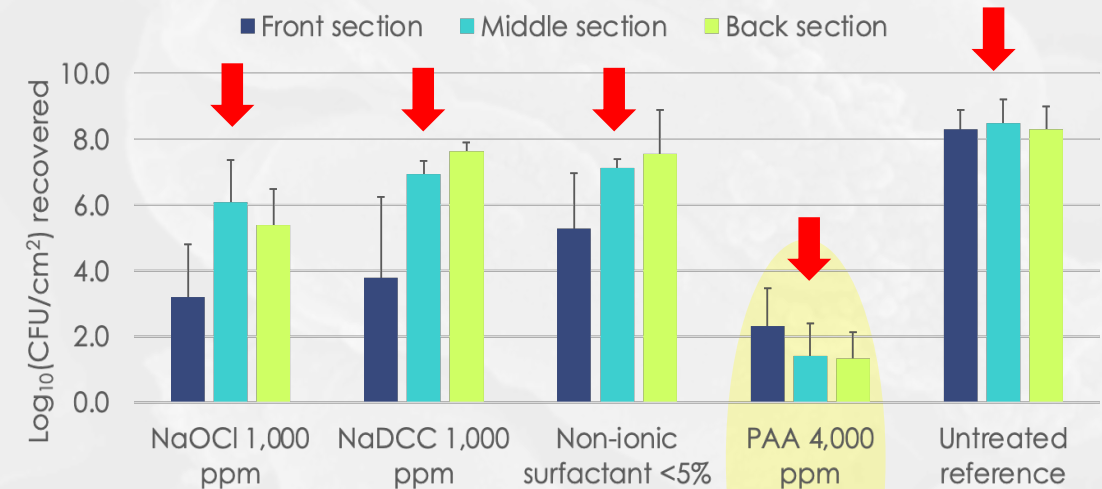
## Disinfection

- Product treatment: 3 x 15 min doses

## Regrowth test

- Drain biofilm bacteria recovered 4 days after product treatment

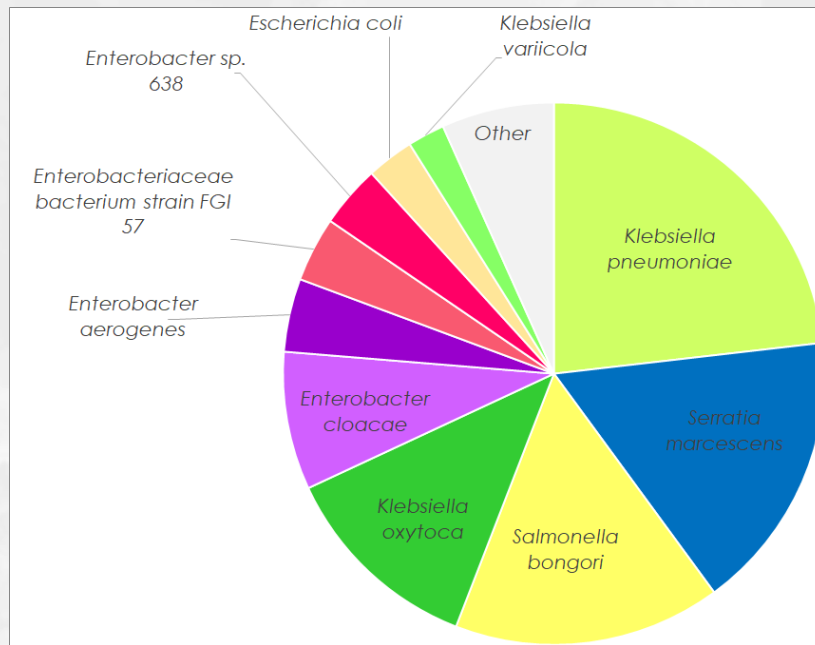
Ledwoch *et al.* *J Hosp Infect* 2020 106;757-764.



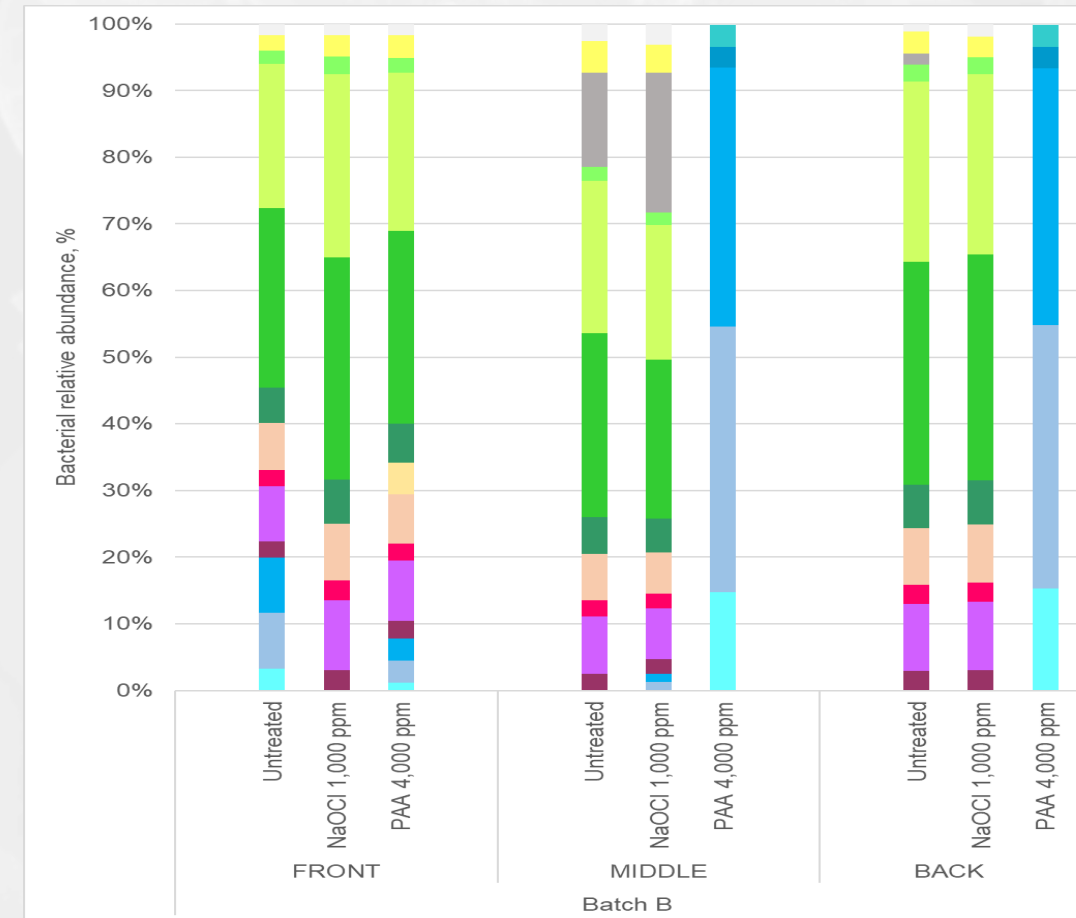
# Efficacy of disinfectants against hydrated biofilms

## Drain biofilms

- Complex microbial communities formed and grown in drain.
- Can migrate to sink and disperse around the sink (2 m)
- Associated with HAIs



Biofilm composition from the small-scale drain model after treatment & regrowth



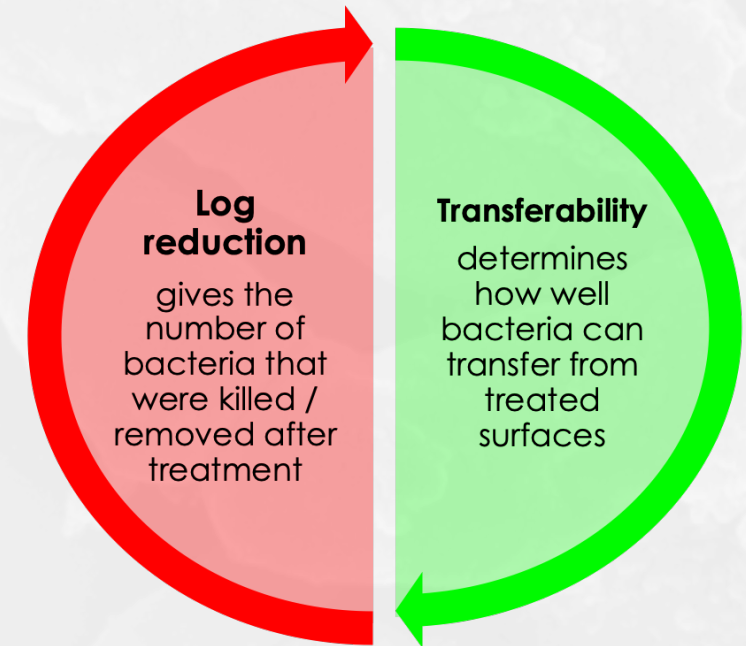
# Efficacy of disinfectants against DSB

Ledwoch *et al.* *J Hosp Infect* 2021;112:31-36.

Treatment: 10 sec wiping (500 g)  
+ 2 min residual time post wiping  
ATM2967-15 "Wiperator test"

Wipe

<i>S. aureus</i> NCTC10788 dry surface biofilm			Log <sub>10</sub> reduction	Direct transferability
			log <sub>10</sub> (CFU/ml)	no of contacts positive for bacterial growth
Treatment	Concentration /power	pH	■ ≥4, ■ <4	■ <2, ■ >2
BZK	< 0.5%	5.41	4.9 ± 2.4	0.0 ± 0.0
ClO <sub>2</sub>	1,000 ppm	4.31	3.0 ± 0.3	31.0 ± 8.7
NaOCl-Ref	1,000 ppm	11.31	4.3 ± 1.3	33.3 ± 3.1
NaOCl-2	1,000 ppm	13.13	2.9 ± 0.4	2.7 ± 4.6
PAA-1	3,500 ppm	8.82	6.3 ± 0.8	0.7 ± 1.2
PAA-2	250 ppm	7.74	6.1 ± 1.5	36.0 ± 0.0
VHP	7% hydrogen peroxide 5% acetic acid 0.4% peracetic acid	n/a	0.8 ± 0.7	36.0 ± 0.0
CAP	P <sub>discharge</sub> = 34.5 W	n/a	0.8 ± 0.0	36.0 ± 0.0
Water	n/a	6.99	2.3 ± 0.4	36.0 ± 0.0
No treatment		n/a	n/a	36.0 ± 0.0



# Efficacy of disinfectants against DSB

Ledwoch et al. ICHE 2021;2:1-7.

<i>S. aureus</i> NCTC10788 dry surface biofilm			Cross transferability (nitrile gloves)	Cross transferability (latex gloves)
			no of contacts positive for bacterial growth	no of contacts positive for bacterial growth
Treatment	Concentration /power	pH	■ <2, ■ >2	■ <2, ■ >2
BZK	< 0.5%	5.41	4.3 ± 1.5	2.7 ± 2.5
ClO <sub>2</sub>	1,000 ppm	4.31	2.3 ± 2.1	1.3 ± 1.2
PAA-1	3,500 ppm	8.82	0.7 ± 0.6	0.0 ± 0.0
PAA-2	250 ppm	7.74	0.0 ± 0.0	0.3 ± 0.6
APG	<1%	5.30	2.3 ± 1.2	4.7 ± 1.5
CP	0.2-0.5%	6.41	1.3 ± 0.6	2.7 ± 2.1
LDAO	0.5%	5.87	0.0 ± 0.0	2.3 ± 3.2
NIS	<5%	7.05	2.0 ± 2.0	2.0 ± 2.6
Water	n/a	6.99	3.0 ± 1.0	6.7 ± 1.5
No treatment		n/a	10 ± 3.0	7.0 ± 1.7

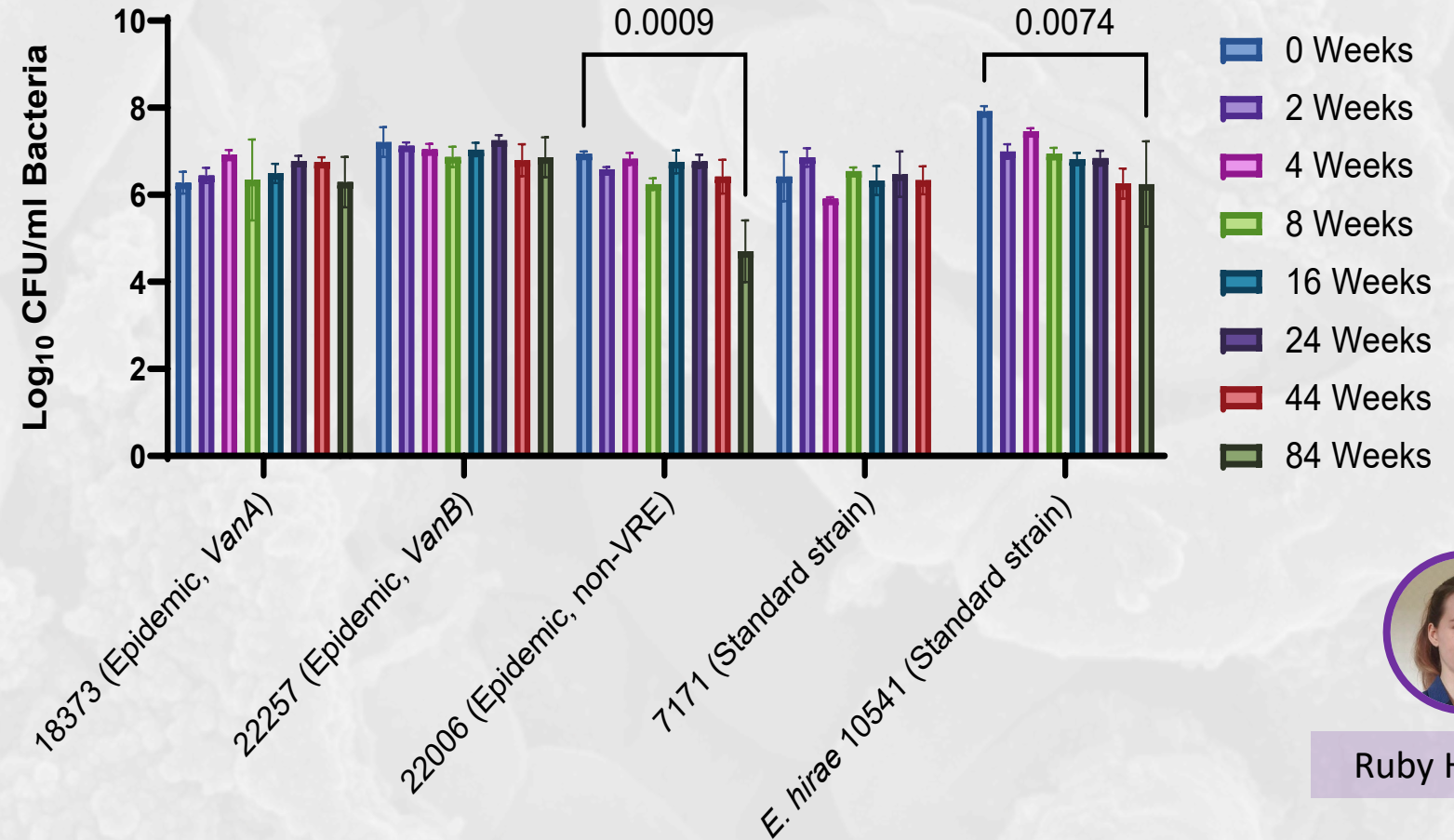
Treatment: 10 sec wiping (500 g)  
+ 2 min residual time post wiping

- High transfer rates using gloves following the use of surfactants.
- Type of gloves matter with latex gloves performing worse than nitrile ones.

# Persistence of DSB on surfaces

- *Enterococcus faecium* DSB & *E. hirae* on stainless steel disks

- > 84 weeks (20°C; 55% RH)
- >6 Log<sub>10</sub> recovered



Ruby Harsent

# Clinical implication of DSB?

Otter *et al. Am J Infect Control* 2013;41:S6-11

Mitchell *et al. Infect Dis Health* 2023;28:290-7.

American Journal of Infection Control 41 (2013) S6-S11

Contents lists available at ScienceDirect

**American Journal of Infection Control**

journal homepage: [www.ajicjournal.org](http://www.ajicjournal.org)

Original research article

**Evidence that contaminated surfaces contribute to the transmission of hospital pathogens and an overview of strategies to address contaminated surfaces in hospital settings**

Jonathan A. Otter PhD<sup>a,b,\*</sup>, Saber Yezli PhD<sup>b</sup>, James A.G. Salkeld BSc<sup>b</sup>, Gary L. French MD, FRCPath<sup>a</sup>

<sup>a</sup>Centre for Clinical Infection and Diagnostics Research (CIDR), Department of Infectious Diseases, King's College London & Guy's and St. Thomas' NHS Foundation Trust, London, UK  
<sup>b</sup>Bioquell, Andover, Hampshire, UK

Infection, Disease & Health xxx (xxxx) xxx

Available online at [www.sciencedirect.com](http://www.sciencedirect.com)

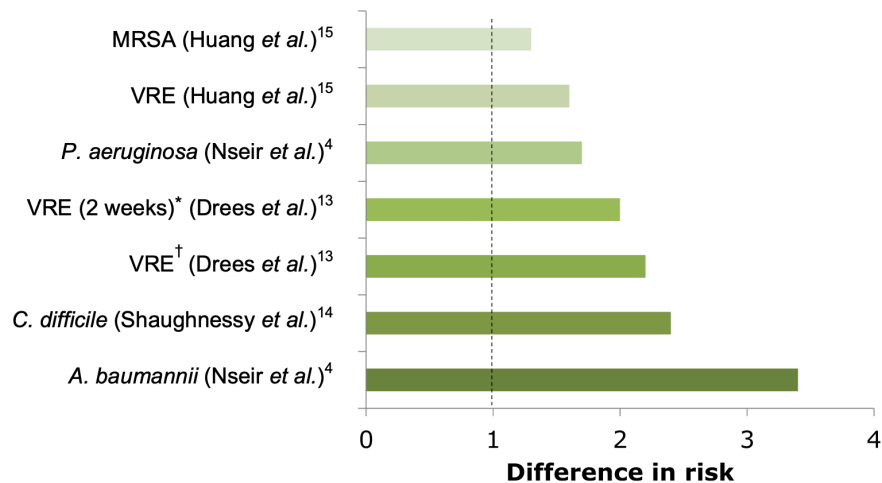
**ScienceDirect**

journal homepage: <http://www.journals.elsevier.com/infection-disease-and-health/>

Review

**Risk of organism acquisition from prior room occupants: An updated systematic review**

Brett G. Mitchell<sup>a,b,c,d,\*</sup>, Julee McDonagh<sup>e,f</sup>, Stephanie J. Dancer<sup>g</sup>, Sindi Ford<sup>h,i</sup>, Jenny Sim<sup>j,k,l,m</sup>, Bismi Thottiyil Sultanmuhammed Abdul Khadar<sup>d,k</sup>, Philip L. Russo<sup>b,n,o</sup>, Jean-Yves Maillard<sup>p</sup>, Helen Rawson<sup>q</sup>, Katrina Browne<sup>a,b</sup>, Martin Kiernan<sup>b,r</sup>



**Fig 1.** Chart showing the increased risk associated with the prior room occupant. The figures of difference in risk are unadjusted based on raw data. Several of the studies included adjusted measures of risk, but these were not included because of differences in study design. \* Any patient infected or colonized with VRE in the two weeks prior to admission. † The immediate prior room occupant was known to be infected or colonized with VRE.

- The risk of pathogen acquisition remains high; pooled acquisition odds ratio for all the pathogens was 2.45 (95% CI: 1.53-3.93]
- Support the need for a clean environment to reduce the risk of healthcare associated infections.

# Biofilms susceptibility to disinfectants

	Antibiotics	Biocides	
	Bacteriostatic	Bactericidal	
Planktonic suspension	MIC determination	Suspension test	
Planktonic dried on surfaces		Carrier test	Product* test (Wiping + transfer)
Hydrated biofilms	MBEC determination	MBIC/MBEC determination	-
		Several US standards	-
		<i>S. aureus</i> / <i>Ps. aeruginosa</i>	-
Dry surface biofilms		-	-

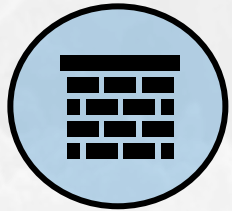
Susceptibility

\* Product: formulation and delivery system

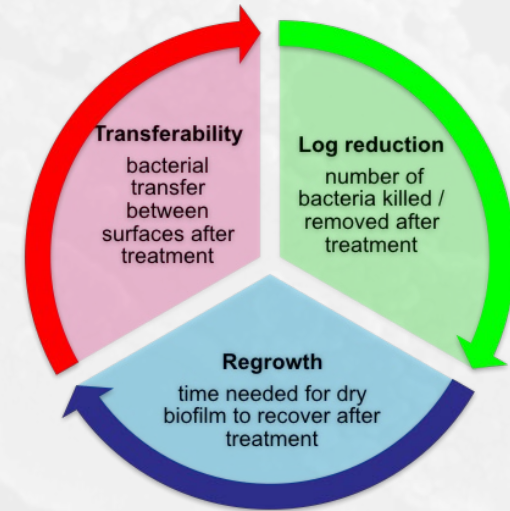
# Biofilms susceptibility to disinfectants



- No “wet” standard biofilm test protocol in Europe
  - Product claim? Ad hoc testing
  - Knowing the end points: **Reduction in viability + regrowth**
  - Review data but need to understand protocol



- No dry surface biofilm standard test protocol
  - Product claim? Ad hoc testing
  - Knowing the end points – **Reduction in viability + transfer**
  - Review data but need to understand protocol



# Acknowledgments



Dr Katrina Duggan



Dr Katarzyna Ledwoch



Dr Isabella Centeleghe



Dr Ruby Harsent

